

## Hematological Characteristics of the Cultured Eel, *Anguilla japonica* by Vibration Stress

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The purpose of this study is to determine the effects of vibration on primary (e.g. plasma cortisol), secondary (e.g. plasma glucose, aspartate aminotransferase (AST), alanine aminotransferase (ALT), Na<sup>+</sup>, K<sup>+</sup> and Cl<sup>-</sup> and tertiary (e.g. mortality) stress responses in cultured eel, *Anguilla japonica*. For this purpose, three groups (including one control group and two stress groups) were set up. The control group was made exposed to vibration corresponding to 48 decibel (dB, V) (produced using electric vibrators) for 15 minutes per hour every day, and the two stress groups was made exposed to vibration corresponding to 58 and 68 dB (V) (produced using the same electric vibrators), equally, for 15 minutes per hour every day. Blood was sampled at day 0 (before starting vibration stress tests, BS), and days 1, 3, 5, 7, 9 and 11 (after starting vibration stress tests). As a result, plasma cortisol showed trend to continuously rise by consecutive stress from 4.1±0.1 ng/ml in BS. In 48 dB group (control), cortisol showed the highest level with 7.6±0.9 ng/ml after 7 days ( $p<0.05$ ), but at 9 and 11 days was not significantly compared with BS level. In 58 dB group, the cortisol showed the highest level with 43.1±4.8 ng/ml after 1st day. Cortisol of 68 dB group increased significantly during the experimental period (14.4±2.3~32.0±5.7 ng/ml) ( $p<0.05$ ). In 58, and 68 dB groups during the experimental period differed significantly compared to 48 dB group ( $p<0.05$ ). Glucose in 48 dB were increased from 42.0±5.7 (BS) to 52.5±2.1 (1 day), the level was not significantly from 1 to 11 days. Glucose in 58 and 68 dB groups was increased significantly than BS during experimental period ( $p<0.05$ ). K<sup>+</sup> in 68 dB increased significantly ( $p<0.05$ ) from 2.3±0.2 mEq/l (BS) to 3.3±0.5 mEq/l at 5 days. In 48 and 58 dB groups during the experimental period differed significantly ( $p<0.05$ ). Na<sup>+</sup> and Cl<sup>-</sup> levels were not differed significantly during the experimental period. AST and ALT in 58 and 68 dB groups showed trend to continuously rise by consecutive stress. At 7 and 9 days in AST, between 48, 58 and 68 dB groups differed significantly ( $p<0.05$ ). In 48, 58 and 68 dB groups at 1 day, blood hematocrit increased significantly higher than BS. The 11 days after vibration stress, the mortality in 48, 58 and 68 dB groups was 1.1, 5.1 and 5.8%, respectively. The present results have shown that *A. japonica* exhibited "typical" physiological responses when exposed to chronic vibration stress. These data suggested that chronic vibration stress caused substantial stress in the fish; especially the persisting elevated plasma AST and ALT levels observed would be expected to adverse effect. In conclusion, chronic vibration stress could greatly affects the hematological characteristics in *A. japonica*.

**Keywords:** Eel, *Anguilla japonica*, Vibration stress, Hematological character

### Introduction

*A. japonica* distributes widely in eastern Asia, mainly in Korea, China, and Japan freshwater system. And it is one of the important

indoors culture species in Korea. On the average, *A. japonica* farm mainly locates in a quiet place with good water condition.

Recently a lot of vibration arose from the development of industrialization and urbanization, roadwork and industrial establish-

ment, this has made serious harmful effect on nearby aquaculture farm (Lee et al., 2006; Lee et al., 2007; Lee and Hur, 2004; Hur et al., 2009; Hur and Lee, 2010; Hur et al., 2015). These construction work mainly started from spring and ended in autumn so can affect the neighboring *A. japonica* farm obviously. Now there was report about the bad influence of vibration on soft-shelled turtle, *Pelodiscus sinensis*, catfish, *Silurus asotus* and *A. japonica* (Lee et al., 2007; Lee and Hur, 2004; Hur et al., 2009; Hur and Lee, 2010; Park and Yoon, 2017), however there is little research on *A. japonica*.

The total production of cultured freshwater fish was 36,160 M/T in Korea in 2017, in which the production of *A. japonica* accounted for 31.0%, about 11,067 M/T (Korean Statistical Information Service, 2017). The production of cultured *A. japonica* has increased from 8,021 M/T (2010) to 9,904 M/T (2016). Compared to other freshwater fish, *A. japonica* culturing has been increasing gradually, as well as the consumption.

Accordingly there is little report on the hematological characteristics to the vibration stress in culture environment domestic and overseas, there is a need to do some research in this field. This study has investigated a short period index for the stress responses: the primary (plasma cortisol), secondary (plasma glucose, AST, ALT,  $\text{Na}^+$ ,  $\text{K}^+$  and  $\text{Cl}^-$ ), and tertiary (mortality).

## Materials and Methods

*A. japonica* (mean length:  $22.9 \pm 0.9$  cm, mean weight:  $295.5 \pm 65.5$  g,  $\pm$  SD,  $n=66$ ) obtained from research station, Kunsan National University, Korea. They were habituated in a lab aquarium tank for 2 weeks before used in the study. The *A. japonica* ( $n=450$ ) were distributed in 9 tanks (2 ton, FRP, raceway style) by experimental scheme, and an electric vibrator was attached to one end-side of tank outer.

The non-stress group (control) of 48 dB (V), stress groups of 58 and 68 dB (V) from an electric vibrator were turned on for 15 min per one hour during 11 days. In order to exclude other stresses, the lab was constantly kept dark, with no other disturbances, and the water temperature was automatically regulated from 24.5 to 25.5°C using a temperature control system. The *A. japonica* in the non-stressed group ( $n=50$ ) were maintained under the same environmental conditions excluding vibration for 11 days.

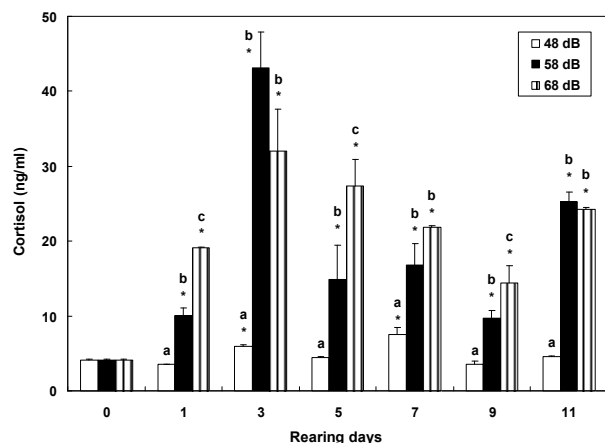
Blood was sampled during 1 min without anesthesia from the caudal blood vessel complex using heparinized syringes at day 0 (before start of the experiment, BE), and at 1, 3, 5, 7, 9, and 11 days after vibration stress.

Hematocrit, red blood cells (RBC) and hemoglobin were analyzed

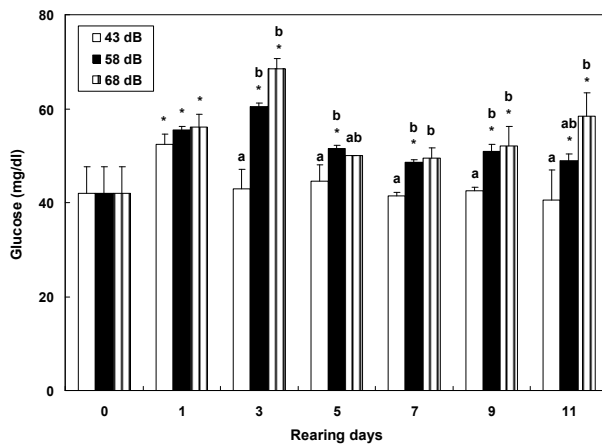
immediately using an automatic blood analyzer (Excell 500, Danam Electronics, Dallas, TX, USA). Blood samples were kept in 2-ml vacuum containers treated with sodium fluoride/potassium oxalate (Vacutainer, UK) and in 1.5-ml polypropylene microcentrifuge tubes held on ice for less than 5 min before centrifugation at 5,600 g for 5 min. Plasma was then collected and stored in a deep freeze (CLN-500 UW Nihon Freezer; Nihon Co., Japan) at  $-70^\circ\text{C}$  until analysis.

Plasma cortisol concentration was determined in 50- $\mu\text{l}$  samples using radioimmunoassay kits (Coat-A-Count TKCO Cortisol RIA Kit; DPC, USA). Mixtures of samples in 100 ml of antiserum were incubated for 45 min at  $37^\circ\text{C}$ , and then, 1,000 ml of separation reagent was added. The mixtures were placed in a refrigerator at  $4^\circ\text{C}$  for 15 min and then centrifuged at 1,200 g for 15 min. The supernatant was assayed for gamma radiation using an automatic gamma counter (Cobra II; Packard Co., USA). Plasma glucose, AST (aspartate aminotransferase) and ALT (alanine aminotransferase),  $\text{Na}^+$ ,  $\text{K}^+$  and  $\text{Cl}^-$  were analyzed using an automatic chemistry analyzer (Hitachi 7180, Hitachi, Japan).

Experiment was performed in triplicate and results were reported as means  $\pm$  SD ( $n=5$ ) unless otherwise stated. Data were analyzed by a one-way ANOVA in the SPSS (Statistical Package for Social Sciences) statistical package. Means were separated by using Duncan's multiple range test and were considered significantly different if  $p < 0.05$ .



**Fig. 1.** Variations of plasma cortisol of *A. japonica* to the vibration stress for 11 days. Values are means of experiments run on two occasions ( $\pm$  SD,  $n=5$ ). Significant differences from control values are indicated by asterisks ( $*p < 0.05$ ). Shared alphabetic letters on shaded bars indicates significant differences between three groups at equivalent days (Duncan's multiple range test  $p > 0.05$ ).



**Fig. 2.** Variations of plasma glucose of *A. japonica* to the vibration stress for 11 days. Values are means of experiments run on two occasions ( $\pm$  SD,  $n=5$ ). Significant differences from control values are indicated by asterisks ( $*p<0.05$ ). Shared alphabetic letters on shaded bars indicates significant differences between three groups at equivalent days (Duncan's multiple range test  $p>0.05$ ).

**Table 1.** Effects of the vibration stress on plasma  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Cl}^-$ , AST and ALT of *A. japonica* for 11 days

Items	Rearing days	Experimental groups		
		48 dB	58 dB	68 dB
$\text{Na}^+$ (mEq/l)	0	151.0 $\pm$ 1.0	151.0 $\pm$ 1.0	151.0 $\pm$ 1.0
	1	151.0 $\pm$ 0.0	144.7 $\pm$ 16.3	155.7 $\pm$ 4.7
	3	154.7 $\pm$ 3.1	154.0 $\pm$ 0.0	153.3 $\pm$ 2.3
	5	154.0 $\pm$ 1.7	151.0 $\pm$ 3.6	155.7 $\pm$ 2.3
	7	150.3 $\pm$ 1.5	150.0 $\pm$ 5.3	153.3 $\pm$ 6.7
	9	151.3 $\pm$ 5.0	153.7 $\pm$ 3.5	152.7 $\pm$ 5.9
	11	155.0 $\pm$ 2.0 <sup>ab</sup>	155.7 $\pm$ 1.5 <sup>b</sup>	151.3 $\pm$ 2.3 <sup>a</sup>
$\text{K}^+$ (mEq/l)	0	2.3 $\pm$ 0.2	2.3 $\pm$ 0.2	2.3 $\pm$ 0.2
	1	2.9 $\pm$ 0.4 <sup>b</sup>	2.2 $\pm$ 0.2 <sup>a</sup>	2.2 $\pm$ 0.3 <sup>a</sup>
	3	2.6 $\pm$ 0.2	2.7 $\pm$ 0.3	2.9 $\pm$ 0.4
	5	2.7 $\pm$ 0.2 <sup>ab</sup>	3.3 $\pm$ 0.6 <sup>a</sup>	3.3 $\pm$ 0.5 <sup>*b</sup>
	7	2.6 $\pm$ 0.2	2.5 $\pm$ 0.5	2.6 $\pm$ 0.4
	9	2.6 $\pm$ 0.5	2.5 $\pm$ 0.6	2.3 $\pm$ 0.3
	11	2.6 $\pm$ 0.2	2.6 $\pm$ 0.3	2.6 $\pm$ 0.2
$\text{Cl}^-$ (mEq/l)	0	113.0 $\pm$ 6.1	113.0 $\pm$ 6.1	113.0 $\pm$ 6.1
	1	114.0 $\pm$ 4.6	113.3 $\pm$ 11.2	116.0 $\pm$ 5.6
	3	111.3 $\pm$ 2.5	116.0 $\pm$ 2.6	117.0 $\pm$ 6.9

**Table 1.** Effects of the vibration stress on plasma  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Cl}^-$ , AST and ALT of *A. japonica* for 11 days (Continued)

Items	Rearing days	Experimental groups		
		48 dB	58 dB	68 dB
$\text{Cl}^-$ (mEq/l)	5	119.7 $\pm$ 5.9	114.3 $\pm$ 7.0	118.3 $\pm$ 2.5
	7	106.7 $\pm$ 2.1	114.0 $\pm$ 11.5	116.3 $\pm$ 4.7
	9	115.0 $\pm$ 7.0	116.7 $\pm$ 5.5	105.7 $\pm$ 8.1
	11	112.3 $\pm$ 0.6 <sup>ab</sup>	117.3 $\pm$ 3.1 <sup>b</sup>	110.0 $\pm$ 3.5 <sup>a</sup>
AST (IU/l)	0	25.5 $\pm$ 0.7	25.5 $\pm$ 0.7	25.5 $\pm$ 0.7
	1	30.0 $\pm$ 1.4 <sup>ab</sup>	30.5 $\pm$ 0.7 <sup>b</sup>	25.5 $\pm$ 2.1 <sup>a</sup>
	3	32.0 $\pm$ 2.8 <sup>*</sup>	36.0 $\pm$ 2.8 <sup>*</sup>	33.0 $\pm$ 1.4
	5	27.5 $\pm$ 3.5 <sup>a</sup>	35.0 $\pm$ 4.2 <sup>*a</sup>	45.5 $\pm$ 0.7 <sup>*b</sup>
	7	29.5 $\pm$ 2.1 <sup>a</sup>	37.5 $\pm$ 2.1 <sup>*b</sup>	47.5 $\pm$ 2.1 <sup>*c</sup>
	9	25.5 $\pm$ 0.7 <sup>a</sup>	43.0 $\pm$ 4.2 <sup>*b</sup>	49.5 $\pm$ 0.7 <sup>*b</sup>
	11	25.5 $\pm$ 3.5	43.0 $\pm$ 7.1 <sup>*</sup>	39.5 $\pm$ 12.0 <sup>*</sup>
ALT (IU/l)	0	1.7 $\pm$ 0.6	1.7 $\pm$ 0.6	1.7 $\pm$ 0.6
	1	1.0 $\pm$ 0.0	1.0 $\pm$ 0.0	1.3 $\pm$ 0.6
	3	1.3 $\pm$ 0.6 <sup>a</sup>	3.2 $\pm$ 0.7 <sup>*b</sup>	3.3 $\pm$ 0.3 <sup>*b</sup>
	5	1.3 $\pm$ 0.6	2.7 $\pm$ 0.9	2.9 $\pm$ 0.4 <sup>*</sup>
	7	1.7 $\pm$ 0.6 <sup>a</sup>	3.3 $\pm$ 0.7 <sup>*b</sup>	3.0 $\pm$ 0.9 <sup>*ab</sup>
	9	1.0 $\pm$ 0.0 <sup>a</sup>	2.7 $\pm$ 0.6 <sup>b</sup>	3.4 $\pm$ 0.5 <sup>*b</sup>
	11	1.3 $\pm$ 0.6 <sup>a</sup>	3.9 $\pm$ 0.5 <sup>*b</sup>	5.3 $\pm$ 0.7 <sup>*c</sup>

ALT: alanine aminotransferase, AST: aspartate aminotransferase

## Results

Plasma cortisol showed trend to continuously rise by consecutive stress from 4.1 $\pm$ 0.1 ng/ml in BE (Fig. 1). In 48 dB group (control), cortisol showed the highest level with 7.6 $\pm$ 0.9 ng/ml after 7 days ( $p<0.05$ ), but at 9 and 11 days was not significantly compared with BE level. In 58 dB group, the cortisol showed the highest level with 43.1 $\pm$ 4.8 ng/ml after 1st day. Cortisol of 68 dB group increased significantly during the experimental period (14.4 $\pm$ 2.3~32.0 $\pm$ 5.7 ng/ml) ( $p<0.05$ ). In 58, and 68 dB groups during the experimental period differed significantly compared to 48 group ( $p<0.05$ ).

Plasma glucose in 48 dB were increased from 42.0 $\pm$ 5.7 (BE) to 52.5 $\pm$ 2.1 (1 day), the level was not significantly from 1 to 11 days (Fig. 2). Glucose in 58 and 68 dB groups was increased significantly than BE during experimental period ( $p<0.05$ ).

Plasma K<sup>+</sup> in 68 dB increased significantly ( $p < 0.05$ ) from  $2.3 \pm 0.2$  mEq/L (BE) to  $3.3 \pm 0.5$  mEq/L at 5 days (Table 1). In 48 and 58 dB groups during the experimental period differed significantly ( $p < 0.05$ ). Na<sup>+</sup> and Cl<sup>-</sup> levels were not differed significantly during the experimental period.

Plasma AST and ALT in 58 and 68 dB groups showed trend to continuously rise by consecutive stress. At 7 and 9 days in AST, between 48, 58 and 68 dB groups differed significantly ( $p < 0.05$ ).

Table 2 shows the variations of RBC, hematocrit and hemoglobin in plasma to a vibration stress for 11 days. In 48, 58 and 68 dB groups at 1 day, hematocrit was increased significantly higher than BE.

The 11 days after vibration stress, the mortality in 48, 58 and

68 dB groups was 1.1, 5.1 and 5.8%, respectively (no data).

## Discussion

In this study, we knew cortisol as a stress index in blood would increase when received stress. The change of cortisol in all experiments was higher than the control at the first day of experiment when given vibration stress. Cortisol at 7 days was still higher than the control at 48 dB group, but when it came to 9 and 11 days the cortisol got back to normal. However, the cortisol didn't recover until the end of the experiment at 58 and 68 dB groups. During the 11 days fish had accepted considerable affect at the continual vibration stimulus. The content of glucose in blood rose with the

**Table 2.** Effects of the vibration stress on blood hematocrit, hemoglobin and red blood cell of *A. japonica* for 11 days

Items	Rearing days	Experimental groups		
		48 dB	58 dB	68 dB
Hematocrit (%)	0	27.8±2.5	27.8±2.5	27.8±2.5
	1	36.2±3.8*	36.0±3.0*	38.1±4.6*
	3	26.2±3.6	32.9±3.0	32.8±2.8
	5	25.4±0.6	43.9±6.2*	35.9±11.0
	7	24.4±4.9 <sup>a</sup>	46.4±4.2 <sup>*b</sup>	62.2±2.8 <sup>*c</sup>
	9	26.7±3.2 <sup>a</sup>	43.9±4.9 <sup>*b</sup>	37.7±3.3 <sup>*ab</sup>
	11	29.3±2.8 <sup>a</sup>	37.4±2.1 <sup>*b</sup>	33.2±0.1 <sup>ab</sup>
Hemoglobin (g/dl)	0	19.6±1.7	19.6±1.7	19.6±1.7
	1	23.1±3.6	17.9±1.2	19.2±1.1
	3	25.0±0.1 <sup>*b</sup>	19.3±2.4 <sup>a</sup>	20.0±1.2 <sup>a</sup>
	5	20.1±0.4 <sup>ab</sup>	16.6±1.4 <sup>a</sup>	27.5±4.8 <sup>*b</sup>
	7	18.0±0.3 <sup>a</sup>	20.4±1.4 <sup>a</sup>	24.5±1.3 <sup>*b</sup>
	9	19.5±0.2	21.5±1.8	18.7±0.6
	11	20.8±2.1	21.8±1.1	18.6±0.2
Red blood cell (×10 <sup>6</sup> cell/μl)	0	2.3±0.1	2.3±0.1	2.3±0.1
	1	2.8±0.2 <sup>a</sup>	3.7±0.1 <sup>b</sup>	3.2±0.5 <sup>ab</sup>
	3	2.9±0.1 <sup>b</sup>	2.3±0.2 <sup>a</sup>	2.5±0.1 <sup>ab</sup>
	5	3.2±0.4*	5.2±1.5*	5.4±1.5*
	7	2.7±0.1 <sup>a</sup>	6.2±0.9 <sup>*b</sup>	7.4±0.7 <sup>*b</sup>
	9	3.3±0.2 <sup>*a</sup>	4.4±0.1 <sup>*b</sup>	3.1±0.1 <sup>a</sup>
	11	2.6±0.4	2.9±0.1	2.5±0.1

increasing of cortisol was the general reaction to stress (Barton and Iwama, 1991).

In this study glucose rose with the increasing of cortisol for the stress affection, this needed much energy. Olive flounder, *Paralichthys olivaceus*, after once acute stress, the variation of cortisol will get back to normal value of 24 hours ago (Hur, 2002; Lim and Hur, 2018). But in this study the vibration stress repeated every day so we could decide the affection at the continuous trend. Moreover the variation of cortisol and glucose on the first day was generally something like the reaction to stress. And the value of cortisol and glucose rose with the increasing of stress. In this experiment 48 dB group was a common vibration in aquaculture farm. Because the other conditions were different from the aqua farm so the same reaction didn't appear in aqua farm fish, in this study the reaction to stress continued to the 7 days.

Hence, some metabolic levels recovered within 24 hours, but a longer time would be required for normal metabolism to take place after fish have experienced stress. According to the report by Pickering (1993), complete physiological recovery from the stresses studied would take as long as 2 weeks.

Responses to stress-related disturbances in fish are often characterized as primary, secondary and tertiary (Hur, 2002). Primary response is the activation of brain centers that eventually results in the release of cortisol from the steroid-producing cells and of catecholamine from the chromaffin cells of the head kidney. Secondary responses are defined as the subsequent actions and effects of these hormones at blood and tissue level, and may include a disturbance of the hydromineral and metabolic balance. Tertiary responses, exemplified by inhibited growth, hampered reproduction and immune suppression, concern the performance of the organism.

In this study plasma AST and ALT had a trend like this (tertiary response). This result showed AST and ALT increased with experiment processing, so we can know whether there was affection on internal organs or not. AST and ALT are aminotransferases that are distributed in the liver and spleen. Levels are low when animals are healthy, but increase when tissue necrosis is present or animals become sick (Almeida et al., 2002). Davis and Parker (1990) reported that hemodynamic indices of oxygen carrying ability are increased by stress. In this research the value of AST and ALT showed a significant difference between the start and end, so we could know whether there was physiology burden on liver, spleen and other tissue at different vibration. The results of AST and ALT in this study were similar to that in a parallel experiment which conducted on *P. sinensis* (Lee et al., 2007).

Davis and Parker (1990) had reported the hematology index-oxygen transmission ability became high when suffered stress. Blood hematocrit, RBC and hemoglobin were the basic blood factors through which oxygen transformation ability could be estimated and in this study they also showed reaction to stress like other blood items accompany with vibration processing.

The decrease of hemoglobin content and RBC indicated that the oxygen supply and demand was in trouble because of stress. The decrease of hemoglobin content made the respiration quotient induce (Rodrigues et al., 1989), and decrease of hemoglobin and RBC in blood which in charge of oxygen receiving and transformation mean oxygen supply and demand was abnormal (Perry and Reid 1993), energy mobilization problems occurred. In case the stress was weak *A. japonica* would increase hemoglobin and RBC, oxygen supply and energy mobilization to overcome the stress. But when under the continuous stress, *A. japonica* was affected seriously and hemoglobin and RBC decreased for depressing of oxygen consumption. In the case of *P. olivaceus*, after temperature and transportation stress hemoglobin and RBC increased abruptly then got back to the beginning condition quickly (Chang et al., 2001; Hur et al., 2007, 2008, 2015). That was to say small stress like temperature, transportation and so on could cause blood characteristic disorder but recovered at once, in this study *A. japonica* couldn't adjust to the continuous stress and when this kind of stress acted on the fish it couldn't recover to the normal.

In this study we could see *A. japonica* had received a lot of stress through the analysis of blood index cortisol, glucose, AST, ALT, RBC and hemoglobin. *A. japonica* was bottom teleostei so they were more sensitive to the vibration and other kind of stress. This experiment had measured physiology activity and several variation items, but we still needs to measure more items and histology variation of internal organs and investigate mechanism different physiology activity variation in the future for the determination of physiology exhaustion under different stress.

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