

Molecular Identification and Effects of Temperature on Survival and Growth of Hybrids between *Haliotis gigantea* Gmelin (♀) and *Haliotis discus hannai* Reeve (♂)

Hye Suck An*, Jong Won Han, Hyun-Ju Hwang, Hancheol Jeon, Seung-Hyun Jung, Seonmi Jo, Tae-Young Choi, Young Se Hyun, Ha Yeun Song, Ilson Whang

Genetic Resources Research Department, National Marine Biodiversity Institute of Korea (MABIK), Chungchungnam-do 33662, Korea

Corresponding Author

Hye Suck An
 Genetic Resources Research Department,
 National Marine Biodiversity Institute of
 Korea (MABIK), Chungchungnam-do
 33662, Korea
 E-mail : mgran@mabik.re.kr

Received : September 27, 2017

Revised : October 20, 2017

Accepted : October 26, 2017

In abalones, interspecific hybridization has been suggested as a possible means to increase production and desired traits for the industry. In Korea, *Haliotis gigantea* is considered a species with a larger size and higher temperature tolerance than *H. discus hannai*. However, *H. discus hannai* is considered the most valuable and popular fishery resource due to its better acceptance and higher market prices. Thus, viable interspecific hybrids have been produced by artificial inseminating *H. gigantea* eggs with *H. discus hannai* sperm. However, the reciprocal hybrid cross was not successful. In this study, the hybridity and the growth and thermal tolerance performance of the interspecific hybrids were examined. A combination of various assays revealed maximum growth occurrence at 21°C and the higher growth rate in the hybrids than that of *H. discus hannai* parent. In addition, the growth and survival at high-temperature (28°C) of the hybrids was equivalent to that of the highly tolerant *H. gigantea* parent, suggesting new possibilities to overcome the mass mortality in *H. discus hannai* during high temperature periods of summer season in Korea. Furthermore, the induced interspecific hybrid status was confirmed by the presence of species-specific bands for each parental species of the random amplified polymorphic DNA (RAPD) profiles using universal rice primer (URP), which could be used as species-specific markers to distinguish the hybrids and their parental species.

Keywords: Korean Pacific abalone; Hybrids of *Haliotis gigantea* and *Haliotis discus hannai*; High-temperature tolerance; Growth; Molecular identification of hybrids

Introduction

Abalone are a group of marine gastropod mollusks belonging to the genus *Haliotis* that comprises over 100 species and are distributed throughout of the world in temperate and tropical oceans (Geiger and Poppe, 2000). Among the more than 30 species of commercially available abalone, six species of Pacific abalone have been reported to occur along the Korean coasts, *H. discus hannai* Ino, *H. discus discus* Reeve, *H. madaka* Habe, *H. gigantea* Gmelin, *H. diversicolor supertexta* Reeve, and *H. diversicolor diversicolor* Reeve (Kim et al., 1988; An et al., 2005). Of these, *H. discus hannai* is the most commercially valuable species and a highly sought-after delicacy in Korea. Seedling production and rearing techniques for *H. discus hannai* have been established and its aquaculture has become increasingly popular because of its consistent high

demand and market value (Zhang et al., 2004). The production of this abalone increased drastically from about 1,000 t/y in 2003, reaching 6,941 t in 2011 (wild catch, 162 t and aquaculture, 6,779 t; Korean Statistical Office, 2012). *H. gigantea* is a valued commercial species distributed along the coast of Jeju Island, Korea. However, the production has not reached important levels due to the poor performance. This species is considered a species with a larger size and higher temperature tolerance than *H. discus hannai*. The meat of *H. gigantea* is crisp and tender, and this species possesses excellent disease resistance (Gao et al., 2000). These traits make it a commercial aquaculture abalone species in Korea. *H. discus hannai* and *H. gigantea* are easily distinguished by their surface features and by the color of their shells.

Currently, the production of intra- and interspecific hybrids is an alternative to conventional selective breeding methods of econom-

ically important aquaculture species (Elliott, 2000). Hybridization has been successfully obtained in many marine species (Bartley et al., 2001; Hulata, 2001). In worldwide abalone aquaculture, more than 50 crosses between sympatric and/or allopatric species have been reported to be successfully produced and most of the hybrid abalone evaluated have shown faster growth, high survival rate, particular adaptations to cultured conditions and desirable market qualities (Lafarga-de la Cruz and Gallardo-Escárate, 2010). Thus, hybridization between *H. discus hannai* and *H. gigantea* has been suggested as a possible method to increase growth rate and adaptability to environmental conditions. Laboratory crosses of *H. discus discus* and *H. gigantea* have demonstrated that F1 hybrids can be produced with a low fertilization rate (An et al., 2007; Ahmed et al., 2008). Genetic identification and biological characterization of hybridized individuals are the first step for developing conservation and management strategies and improving the aquaculture.

Advances in molecular genetics in recent years have provided several types of genetic markers to be used to identify inter- and intraspecies levels and hybridization in aquatic animals (Liu and Cordes, 2004; Choi et al., 2012). Among them, the random amplified polymorphic DNA-polymerase chain reaction (RAPD-PCR) technique is a relatively simple and cheap method capable of differentiating taxa without the need to know their genomes (Williams et al., 1990). RAPD markers have been used to study species identification and genetic relationships for many other marine organisms including abalone (Klinbunga et al., 2004). URPs (universal rice primers) were developed from repetitive sequence of rice genome (Kang et al., 2002). URP-PCR technique is a useful tool for the characterization and grouping most eukaryotic or prokaryotic genomes, especially at inter- and intraspecies levels (Kang et al., 2002; Jana et al., 2005).

Temperature is a direct and primary environmental factor determining an aquatic organism's activity and therefore mobile species including the abalones show different behavioral responses. Hence, when culturing a species, best performance is obtained when temperatures are optimal (Díaz et al., 2000). Furthermore, high temperature during summer is apparently a major constraint for abalone farming in Korea. Therefore, the thermal threat has become one of the major challenges for abalone farming in Korea. An efficient and moderate method that can precisely predict abalone thermal performance would be greatly beneficial in breeding projects. Thus, ascertaining effects of temperature on survival and growth of the hybrids is very important due its pivotal role for the abalone physiology.

The main objective of this study was to confirm the hybridity of the hybrids between *H. gigantea* (♀) and *H. discus hannai* (♂) using URP-PCR approach and to evaluate effects of temperature on growth and survival of the hybrids in order to assess the ability to adapt to different thermal regimes. These data will be useful for developing conservation and management strategies and improving the aquaculture industry of abalone fisheries in Korea.

Materials and Methods

1. Samples

The broodstock of *H. gigantea* (G) and *H. discus hannai* (D) were originally from Jeju Island and Pohang, Korea respectively. Two species were collected between February and May of 1999 and 2000.

Hybrids between *H. gigantea* and *H. discus hannai* was produced (GD, *H. gigantea* ♀ × *H. discus hannai* ♂) and two purebred controls (DD, *H. discus hannai* ♀ × *H. discus hannai* ♂ and GG, *H. gigantea* ♀ × *H. gigantea* ♂) in 2000 at the Shellfish Genetic and Breeding Research Center, Jeju Fisheries Research Institute, National Fisheries Research and Development Institute, Jeju Island, Korea. After artificial insemination, the mantle muscle of parental abalones were sampled and stored in 99% ethanol for DNA extraction. The hybrids and progeny of purebred were reared at tanks separately to prevent intermix. Young abalones from each genotype were raised in multiple tanks per genotype and fed microalgae that grew on the surfaces of the collectors until their shell length was over 6 mm. After that, they were kept in a cage made from netting with a mesh opening size of 2 mm and fed marine algae.

For molecular identification of the hybrid, parental abalones and live juvenile abalones of GD cross, which were randomly sampled at four months of age, were used for RAPD analysis.

For growth and thermal tolerance analysis, live juvenile abalones of each cross were sampled at seven months of age (approximately 1.05 cm shell length and 0.13 g body weight) and reared at each temperature.

2. DNA extraction

Total genomic DNA was extracted from mantle musculature samples of the juvenile abalone of GD cross and parental abalones. The removed musculatures were minced and lysed by TNES-Urea buffer (6 or 8 M urea: 10 mM Tris- HCL, pH 7.5: 125 mM NaCl:

10 mM EDTA: 1% SDS) (Asahida et al., 1996). The genomic DNA was precipitated from the lysate with 100% ethanol. Following an ethanol wash the DNA was solubilized in water (or 8 mM NaOH). DNA was diluted with water to a final concentration of 20 ng/μl.

3. Genetic identification of the hybridity

Primers and PCR conditions are adopted from Kang et al. (1997). Sequences of URP primers are listed in Table 1. Amplification of abalone genomic DNA using URP primers was performed in a PTC-100 thermal cycler (MJ Research, USA). PCR reactions were done in 50 μl volumes. Each reaction contained 5 μl of *10X EX TaqTM* buffer (Takara, Shiga, Japan), 0.2 mM each dNTP, 2 μl of each primer (100 ng), 2.5 units of *EX TaqTM* polymerase (Takara), and 5 μl genomic DNA. The temperature regime used for the amplification was 35 repetitions of a three-step cycle consisting of denaturation at 95°C for 1 min, annealing at 55°C for 1 min, and extension at 72°C for 2 min. Denaturation for the first cycle was 4 min at 95°C and extension for the final cycle was 7 min at 72°C. Fifteen μl URP-PCR products were electrophoresed in 2% agarose gel with TAE buffer. DNA fragments in the gel were visualized by staining with ethidium bromide and photographed under UV transilluminator.

The primers that produced good amplification (clearly defined bands that differentiated between parental abalones) were used in the next amplification step. At this stage, primers were considered as informative and reliable for identification of the hybrid when they produced a specific amplicon both in all individuals of each parental abalone and in all individuals of hybrid.

4. Effects of temperature on survival and growth of the hybrids

In order to investigate effects of temperature on growth and survival of the hybrid and their purebred abalones, the experiments carried out indoors in a flow through system consisting of 30 tanks (35 cm in width × 28.5 cm in length × 23 cm in height) that each served as water baths. A thermoregulator was connected to one extreme for cooling and a 1,000 W heater was placed in the other extreme to keep a thermal temperature between 7~28°C. The vertical thermal stratification was eliminated by use of air stones placed in each tank.

The experiment had two replicate designs with five temperatures [ambient (16.7~28.1°C), 7, 14, 21, and 28°C] per three crosses (GD, DD and GG). Overall 30 trials were conducted. Each trial, 30

Table 1. List of 12 URP primers used in PCR

Primers	Sequences (5' - 3')	GC content (%)	T _m (°C)
URP1F	ATCCAAGGTCCGAGACAACC	50	55
URP2F	GTGTGCGATCAGTTGCTGGG		
URP2R	CCCAGCAACTGATCGCACAC		
URP4R	AGGACTCGATAACAGGCTCC		
URP6R	GGCAAGCTGGTGGGAGGTAC		
URP9F	ATGTGTGCGATCAGTTGCTG		
URP13R	TACATCGCAAGTGACACAGG		
URP17R	AATGTGGGCAAGCTGGTGGT		
URP25F	GATGTGTTCTTGGAGCCTGT		
URP30F	GGACAAGAAGAGGATGTGGA		
URP32F	TACACGTCTCGATCTACAGG		
URP38F	AAGAGGCATTCTACCACCAC		

juvenile abalones of each cross (mean 1.05 cm initial shell length and mean 0.13 g weight) were allocated to each tank. Abalones were exposed to each temperature for 136 days (14-May, 2001~27-Sep, 2001). Feed was offered twice a week. All treatments were offered the same amount of feed as the tanks that had the largest biomass, thus ensuring that ration size did not further increase the growth of faster growing abalone. Mortalities were checked daily during the experiments and recorded and removed. Every month after the start of the experiments, individual weight and length of all abalones from each tank were measured.

Data were analyzed using SYSTAT 10 statistics software. Differences in survival and growth of each phenotype were assessed by ANOVA (followed by Duncan's multiple range test). Significance was set at $p < 0.05$.

Results

1. Genetic evidence for hybridisation

The RAPD-PCR technique can produce non-reproducible amplification products (Harry et al., 1998). Thus, reactions were performed following strict protocol with standardized conditions. Also, all amplification reactions were carried out, at least, three times in

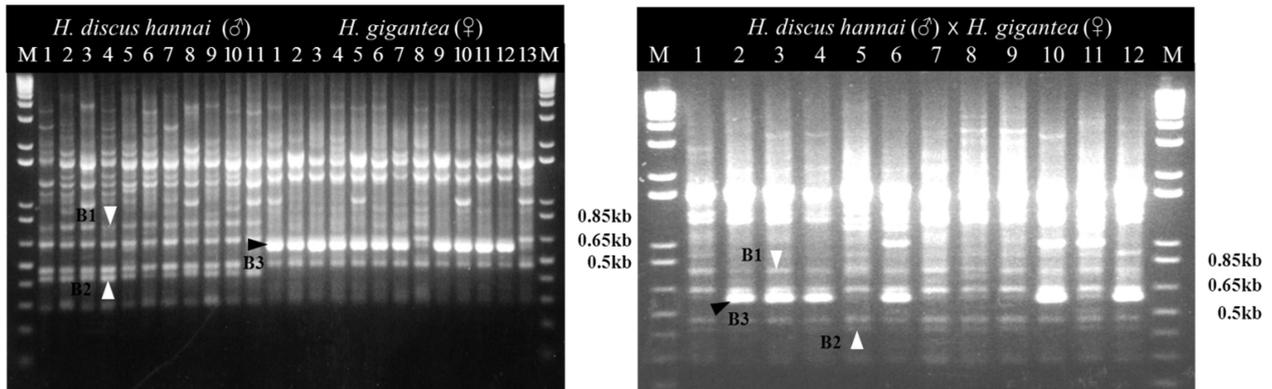


Fig. 1. RAPD band pattern for the male *H. discus hannai*; the female *H. gigantea* (above) and their hybrid (below) generated by URP2R primer. PCR products were resolved by agarose gel (2%) and visualized by staining with ethidium bromide. M: 1 kb ladder (Promega, USA), B1 and B2: two specific amplicons in the male *H. discus hannai* only, B3: a specific amplicon in the female *H. gigantea* only.

order to score reproducible bands clearly.

Hybrid identification was performed by 12 different URP primers (Table 1). They were used as template to assess the wide distribution of the URP nucleotide motif sequences in the parental abalone species genome. The number of amplified bands detected varied, depending on the primers, parental species and individuals. Out of these 12 primers screened, five primers generated reproducible profiles (URP1F, URP2F, URP2R, URP6R and URP30F) and showed different profiles between two parental species. The rest seven primers did not amplify at all or produced highly inconsistent amplification products from the same individuals and were excluded from further analysis. Among 5 primers URP2R only produced both unambiguous patterns of their parents in the hybrid. The gel pictures showing the banding patterns of RAPD profiles generated by URP2R are shown in Fig. 1. The male *H. discus hannai* had two specific amplicons B1 and B2 only. The female *H. gigantea* had a specific amplicon B3 only. The hybrid had a heterotic combination of all parental species specific amplicons B1, B2 and B3.

2. Effects of temperature on survival and growth of hybrids

In order to examine the effect of temperature on survival of hybrids, observations on the survival rate of juvenile abalones of DD, GG and GD crosses were studied at ambient (16.7~28.1°C), 7, 14, 21, and 28°C, respectively. Overall, optimal culturing temperatures of ambient, 14 or 21°C had no effect on survival of

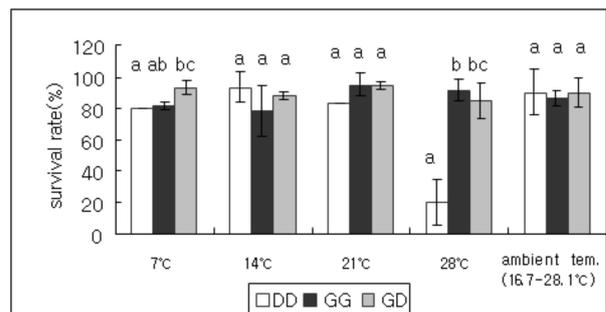


Fig. 2. Mean survival rate of juvenile abalones of DD, GG and GD crosses, cultured at either ambient temperature (16.7~28.1°C), 7, 14, 21 or 28°C during a 136 days trial. Error bars indicate error of the mean of two replicate tanks. DD, the female *H. discus hannai* × the male *H. discus hannai*; GG, the female *H. gigantea* × the male *H. gigantea*; GD, the hybrid between the male *H. discus hannai* and the female *H. gigantea*.

juvenile abalones of three crosses. However, the survivals at low and high-temperature (7 and 28°C) of the hybrids were equivalent to that of the highly tolerant *H. gigantea* parent (Fig. 2). This result may reflect a heterotic effect on survival at low and high water temperatures in the case of the hybrid between the two species compared with those of juvenile abalones of DD cross. Especially, the survival rate of juvenile abalones of DD cross cultured at 28°C was rapidly reduced after 3 months exposure (Fig. 3).

In order to examine the effect of temperature on the growth characteristics of the hybrid, observations on the growth rate of

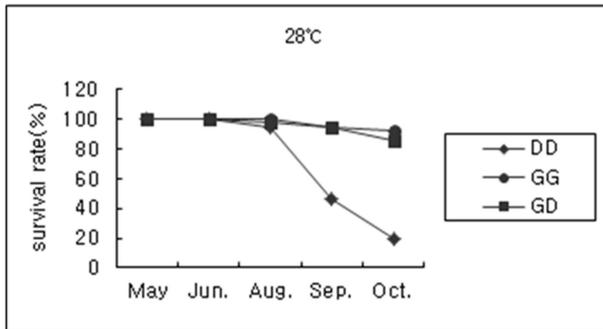


Fig. 3. Mean survival rate of juvenile abalones of DD, GG and GD crosses, cultured at 28°C during a 136 days trial. DD, the female *H. discus hannai* × the male *H. discus hannai*; GG, the female *H. gigantea* × the male *H. gigantea*; GD, the hybrid between the male *H. discus hannai* and the female *H. gigantea*. The survival rate of DD abalones was rapidly reduced after 3 months of exposure.

juvenile abalones of DD, GG and GD crosses were studied at ambient (16.7~28.1°C), 7, 14, 21, and 28°C, respectively. Fig. 4 shows the daily growth in shell length and total weight for juvenile abalones of three crosses. Overall, low culturing temperatures of 7 and 14°C had no effect on the growth rate of juvenile abalones of three crosses. However, at ambient, 21 and 28°C, the values for juvenile abalones of GG cross (154.23±3.26, 169.11±1.43 and 129.22±0.83, respectively) were followed by those for the hybrid (129.61±3.59, 152.25±3.93 and 117.42±4.48, respectively), which was significantly higher than those of juvenile abalones of DD cross (114.70±2.02, 130.03±2.21 and 104.79±1.62, respectively) ($p < 0.05$). Therefore, the growth rates of the hybrid were intermediate between parental species at these relatively high temperatures (Fig. 4, A). Similar tendency was also detected in the daily growth in total weight for juvenile abalones of three crosses except no significant difference between the hybrid and juvenile abalones of DD cross at ambient and 28°C (Fig. 4, B).

Discussion

Abalone hybrid production is difficult and success may reflect genetic affinities between species. A variety of approaches have been used to identify natural or artificial induced hybrids. The verification includes the use of morphological evidences, allozyme and RAPD markers. However, morphological differences of hybrids such as shell characteristics are only minor and need close examination by experienced workers and it is difficult to verify the

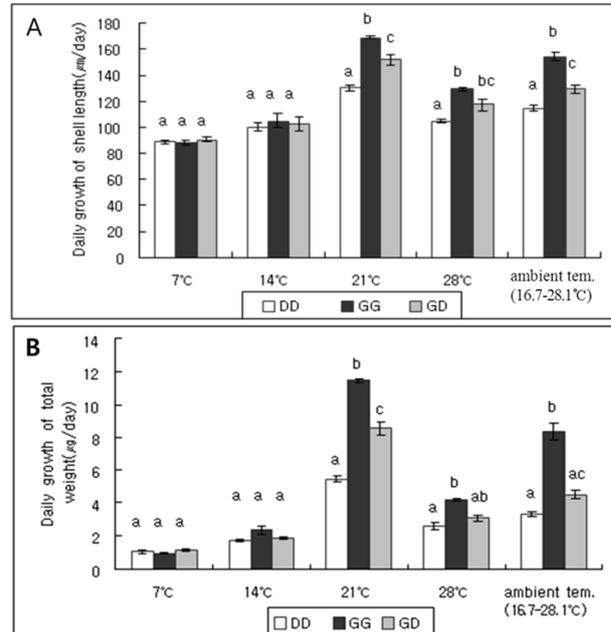


Fig. 4. Daily growth of shell length (A) and total weight (B) in juvenile abalones of DD, GG and GD crosses, cultured at either ambient temperature (16.7~28.1°C), 7, 14, 21 or 28°C during a 136 days trial. Error bars indicate error of the mean of two replicate tanks. DD, the female *H. discus hannai* × the male *H. discus hannai*; GG, the female *H. gigantea* × the male *H. gigantea*; GD, the hybrid between the male *H. discus hannai* and the female *H. gigantea*.

genuine hybrids from interspecific hybridization (Ibarra et al., 2005). Previous studies using allozymes indeed described the genetic difference between the hybrids and their parental species (Ibarra et al., 2005; Ahmed et al., 2008). However, the use of allozyme loci failed to provide a sufficient genetic basis of hybrids, probably due to the less informative nature of allozyme loci. Mutation at the DNA level that causes a replacement of a similarly charged amino acid may not be detected by allozyme electrophoresis, although allozymes represent actual gene products. The information provided by dominant markers yielded by RAPD primers could be useful to facilitate the detection of genetic contributions of different species (Rajora and Rahman, 2003). The main reason for the success of RAPD analysis is the gain of a large number of genetic markers that require small amounts of DNA without the need of the taxa under study. There are many species in which such markers were used, including various abalones (Klinbunga et al., 2004; Marín et al., 2007).

Verification of the hybrid nature in interspecific crossing is

indispensable for genetic improvement of abalones. In this study, we present reliable and highly reproducible procedures for the identification of hybrids between the male *H. discus hannai* and the female *H. gigantea*. *H. discus hannai* and *H. gigantea* are genetically very distinct. In our study, RAPD band profiles of URP2R primer allowed us to clearly distinguish both the parental species and their hybrids. As RAPD is the dominant marker, it has additional advantages with high sensitivity and high reproducibility. RAPD band profiles of the hybrids resulted in the presence of parent-specific bands, which were inherited from both parents. The information yielded by these bands could be sufficient for the identification of interspecific hybrids. Thus, the fact that the hybrid showed the simultaneous presence of the species-specific bands of both parental species indicates that genetic hybridization was accomplished in the cross between the male *H. discus hannai* and the female *H. gigantea*.

The pivotal role of temperature in controlling the rate of metabolic processes is well established and is reflected in survival and growth rates of poikilothermic animals including abalone. Several abalone hybrids have shown positive heterosis (*hybrid vigour*), such as faster growth and higher survival rates than at least one of the parental species (Lafarga-d la Cruz and Gallardo-Escárate, 2011). Our results concur with previous studies, showing that the survival and growth rates in shell length and total weight of juvenile abalone of *H. gigantea* were superior to those of *H. discus hannai* at water temperature 28°C and the survival and growth rates of the hybrid was equivalent to those of the highly tolerant *H. gigantea* parent at water temperature 7 and above 21°C. Similarly, the juvenile hybrid between a female *H. discus hannai* and a male *H. gigantea* also showed growth superior to that of its parental species (Hara, 1992). Furthermore, our result also showed that the survival rate of juvenile abalones of *H. discus hannai* parent was rapidly decreased at high-temperature (28°C). Increasing water temperature can also act as a stressor impacting the immune responses of mollusks, associated with increased susceptibility to disease out-breaks (Dang et al., 2012). Therefore, temperature increase can be a major environmental stressor on marine organisms. When Pacific abalone was exposed to 28°C for 7 days, a dramatic reduction in HSP70 expression occurred. This drop in HSP70 level of juvenile abalones of *H. discus hannai* might be caused by intolerance to long-term exposure at high temperature and this intolerance had caused the breakdown of homeostasis in these juvenile abalones (Liang et al., 2014).

The results of the present study showed the successful confirmation of hybrid status between *H. discus hannai* and *H. gigantea*

using RAPD markers and the early survival and growth traits of these hybrids. Considering that hybridization is particularly useful if the heritability of a trait is low, the intra-species selection may be inefficient (Elliot, 2000). Some underpinning strategic applied research should be conducted to investigate further the hybrid between *H. discus hannai* and *H. gigantea* abalones. From the results of this study, a breeding strategy using hybrids will be available for future abalone culture that is especially applicable to warm temperature water areas. The traits intermediate between parents would be useful for selective breeding material. Furthermore, given that *H. discus hannai* is the most popular and best priced abalone product in the international market, due to the quality of its meat and traditional consumer preferences (Cook and Gordon, 2010), the selection of specific hybrid genotypes could preserve the phenotypic attributes of the meat of *H. discus hannai*, such as flavor, tenderness and texture, as well as the biotechnological solutions to improve fertilization rates of hybrid abalones, given its potential to diversify the Korean abalone industry.

Acknowledgement

This work was funded by a grant from the National Marine Biodiversity Institute of Korea (MABIK), contribution number 2017-M01200. The views expressed herein are those of the authors and do not necessarily reflect the views of MABIK.

References

- Ahmed F, Koike Y, Strussmann CA, Yamasaki I, Yokota M, Watanabe S. 2008. Genetic characterization and gonad development of artificially produced interspecific hybrids of the abalones, *Haliotis discus discus* Reeve, *Haliotis gigantea* Gmelin and *Haliotis madaka* Habe. *Aquacul Res* 39: 532-541.
- An HS, Jee YJ, Han SJ, Kim BL, Kim EM, Park IS. 2007. Induction of a New Hybrid between *Haliotis gigantea* Gmelin (♀) and *H. discus discus* Reeve (♂). *Korean J Genetics* 29: 239-244.
- An HS, Jee YJ, Min KS, Kim BL, Han SJ. 2005. Phylogenetic analysis of six species of Pacific abalone (Haliotidae) based on DNA sequences of 16s rRNA and cytochrome c oxidase subunit I mitochondrial genes. *Mar Biotechnol* 7: 373-380.
- Asahida T, Kobayashi T, Saitoh K, Nakayama I. 1996. Tissue preservation and total DNA extraction from fish stored at ambient temperature using buffers containing high concentration of urea. *Fisher Sci* 62: 727-730.
- Bartley DM, Rana K, Immink AJ. 2001. The use of inter-specific

- hybrids in aquaculture and fisheries. *Rev Fish Biol Fish* 10: 325-337.
- Choi CG, Kim JM. 2012. Detection of *Laminariaceae* Species Based on PCR by Family-specific ITS Primers. *Fish Aquat Sci* 2: 157-162.
- Cook PA, Gordon RH. 2010. World abalone supply, market and pricing. *J Shellfish Res* 29: 569-571.
- Dang VT, Speck P, Benkendorff K. 2012. Influence of elevated temperatures on the immune response of abalone, *Haliotis rubra*. *Fish Shellfish Immunol* 32: 732-740.
- Diaz F, del Rio-Portilla MA, Sierra E, Aguilar M, Re-Araujo AD. 2000. Preferred temperature and critical thermal maxima of red abalone *Haliotis rufescens*. *J Thermal Bio* 25: 257-261.
- Elliott NG. 2000. Genetic improvement programmes in abalone: what is the future? *Aquacult Res* 31: 51-59.
- Gao SS, Wang Q, Wang RB, Li X. 2000. *Abalone*. Liaoning Science and Technology Press, Shenyang, China, 14pp (in Chinese).
- Geiger DL, Poppe GT. 2000. A Conchological Iconography. In *The Family Haliotidae*, Geiger DL and Poppe GT, eds., Conch Books, Hackenheim, Germany, pp 135.
- Hara M. 1992. Breeding of abalone-cross and selection. *Fish Genet Breed Sci* 18: 1-12.
- Hulata G. 2001. Genetic manipulations in aquaculture: a review of stock improvement by classical and modern technologies. *Genetica* 111: 155-173.
- Ibarra AM, Hernandez-Ibarra NK, Cruz P, Perez-Enriquez R, Avila S, Ramirez JL. 2005. Genetic certification of presumed hybrids of blue × red abalone (*Haliotis fulgens* Philippi and *H. rufescens* Swainson). *Aquacult Res* 36: 1-13.
- Jana TK, Singh NK, Koundal KR, Sharma TR. 2005. Genetic differentiation of charcoal rot pathogen, *Macrophomina phaseolina* into specific groups using URP-PCR. *Can J Microbiol* 51: 159-164.
- Kang HW, Go SJ, Eun MY, Chung TY. 1997. Genetic characterization of repetitive sequence derived from Korean red rice. Proceeding of the 8th SABRAO general congress and the Annual Meeting of the Korean, 24-28 September, Seoul, Korea, 421-422.
- Kang HW, Park DS, Go SJ, Eun MY. 2002. Fingerprinting of diverse genomes using PCR with universal rice primers generated from repetitive sequence of Korean weedy rice. *Mol Cells* 13: 281-287.
- Kim Y, Jee YJ, Kim SH, Baik JM, Yang SG. 1988. Interspecific characteristic of abalones in the southern waters of Korea. *Bull Nat Fish Res Dev Agency* 42: 71-80.
- Klinbunga S, Ampayup P, Leelatanawit R, Tassanakajon A, Hirono I, Auki T, Jarayabhand P, Menasveta P. 2004. Species identification of the tropical abalone (*Haliotis asinina*, *Haliotis ovina* and *Haliotis varia*) in Thailand using RAPD and SCAR markers. *J Bioch Mol Biol* 37: 213-222.
- Klinbunga S, Ampayup P, Tassanakajon A, Jarayabhand P, Yoosukh W. 2000. Development of species-specific markers of the tropical oyster (*Crassostrea belcheri*) in Thailand. *Mar Biotechnol* 2: 476-484.
- Korea National Statistical Office. 2012. KOSIS statistical DB. Daejeon, Korea. Available at <http://kostat.go.kr>.
- Lafarga-de la Cruz F, Gallardo-Escarate C. 2011. Interspecies hybrids in *Haliotis*: natural and experimental evidence and its impact on abalone aquaculture. *Rev Aquacult* 3: 74-99.
- Liang S, Luo X, You W, Luo L, Ke C. 2014. The role of hybridization in improving the immune response and thermal tolerance of abalone. *Fish Shellfish Immunol* 39: 69-77.
- Liu ZJ, Cordes JF. 2004. DNA marker technologies and their applications in aquaculture genetics. *Aquaculture* 238: 1-37.
- Marín SA, Haye PA, Marchant S, Winkler FM. 2007. Molecular markers used to analyze species-specific status in abalones with ambiguous morphology. *J Shellfish Res* 3: 833-837.
- Rajora OP, Rahman MH. 2003. Microsatellite DNA and RAPD fingerprinting, identification and genetic relationships of hybrid poplar (*Populus* × *Canadensis*) cultivars. *Theor Appl Genet* 106: 470-477.
- Williams JKG, Kubelik AR, Livak KJ, Rafalsky JA, Tyngey SV. 1990. DNA polymorphisms amplified by arbitrary primers are useful as genetic markers. *Nucleic Acids Res* 18: 6531-6535.
- Zhang H, Que X, Liu X, Xu H. 2004. Abalone mariculture in china. *J Shellfish Res* 23: 947-950.